



UDP-sugars activate P2Y₁₄ receptors to mediate vasoconstriction of the porcine coronary artery

Zainab S.B. Abbas^a, M. Liaque Latif^a, Natalia Dovlatova^{b,1}, Sue C. Fox^{b,1}, Stan Heptinstall^{b,1}, William R. Dunn^a, Vera Ralevic^{a,*}

^a School of Life Sciences, University of Nottingham, Nottingham NG7 2UH, UK

^b School of Medicine, University of Nottingham, Nottingham NG7 2UH, UK

ARTICLE INFO

Keywords:

Coronary artery
P2Y₆ receptor
P2Y₁₄ receptor
UDP-glucose
UDP sugars
MRS2690
Sugar nucleotides

ABSTRACT

Aims: UDP-sugars can act as extracellular signalling molecules, but relatively little is known about their cardiovascular actions. The P2Y₁₄ receptor is a G_{i/o}-coupled receptor which is activated by UDP-glucose and related sugar nucleotides. In this study we sought to investigate whether P2Y₁₄ receptors are functionally expressed in the porcine coronary artery using a selective P2Y₁₄ receptor agonist, MRS2690, and a novel selective P2Y₁₄ receptor antagonist, PPTN (4,7-disubstituted naphthoic acid derivative).

Methods and results: Isometric tension recordings were used to evaluate the effects of UDP-sugars in porcine isolated coronary artery segments. The effects of the P2 receptor antagonists suramin and PPADS, the P2Y₁₄ receptor antagonist PPTN, and the P2Y₆ receptor antagonist MRS2578, were investigated. Measurement of vasodilator-stimulated phosphoprotein (VASP) phosphorylation using flow cytometry was used to assess changes in cAMP levels. UDP-glucose, UDP-glucuronic acid UDP-N-acetylglucosamine (P2Y₁₄ receptor agonists), elicited concentration-dependent contractions of the porcine coronary artery. MRS2690 was a more potent vasoconstrictor than the UDP-sugars. Concentration dependent contractile responses to MRS2690 and UDP-sugars were enhanced in the presence of forskolin (activator of cAMP), where the level of basal tone was maintained by addition of U46619, a thromboxane A₂ mimetic. Contractile responses to MRS2690 were blocked by PPTN, but not by MRS2578. Contractile responses to UDP-glucose were also attenuated by PPTN and suramin, but not by MRS2578. Forskolin-induced VASP-phosphorylation was reduced in porcine coronary arteries exposed to UDP-glucose and MRS2690, consistent with P2Y₁₄ receptor coupling to G_{i/o} proteins and inhibition of adenylyl cyclase activity.

Conclusions: Our data support a role of UDP-sugars as extracellular signalling molecules and show for the first time that they mediate contraction of porcine coronary arteries via P2Y₁₄ receptors.

1. Introduction

Cellular release of purine nucleotides, including UTP and ATP, is promoted by stimuli including hypoxia and inflammation leading to activation of cell-surface P2 receptors [8]. UDP-glucose can also be released from cells to act as an extracellular signalling molecule [32,35,36]. In preliminary studies we showed that UDP-glucose causes constriction of the porcine coronary artery [1–3]. Excessive contraction

of the coronary circulation following hypoxic or inflammatory release of UDP-glucose would exacerbate the detrimental effects of these insults. This led us to investigate the hypothesis that the P2Y₁₄ receptor, a receptor for UDP-glucose and UDP, is expressed in, and mediates the vasomotor role of UDP-sugars in the porcine coronary vasculature. Identification of the receptor(s) involved in contractile responses to UDP-sugars may have therapeutic potential in ischaemic heart disease.

P2Y receptors are G protein-coupled receptors activated by purine,

Abbreviations: BSA, bovine serum albumin; DABCO, 1,4-diazabicyclo[2.2.2]octane; DAPI, 4',6-diamidino-2-phenylindole; ERK, extracellular signal-regulated kinases; FITC, fluorescein isothiocyanate; MAPK, mitogen-activated protein kinases; PPTN, 4-((piperidin-4-yl)-phenyl)-7-(4-(trifluoromethyl)-phenyl)-2-naphthoic acid; RCF, relative centrifugal force; KCl, potassium chloride; MRS2578, N,N'-1,4-butanediylbis[N'-(3-isothiocyanatophenyl)thiourea]; MRS2690, diphosphoric acid 1- α -D-glucopyranosyl ester 2-[(4'-methylthio)uridin-5"-yl] ester disodium salt; NO, nitric oxide; L-NAME, N ω -nitro-L-arginine methyl ester; NTPDase1, nucleotide triphosphate diphosphohydrolase-1; OCT, optimal temperature cutting compound; PAGE, polyacrylamide gel electrophoresis; PBS, phosphate buffered saline; SDS, sodium dodecyl sulphate; SNP, sodium nitroprusside; TBST, Tris-buffered saline and Tween 20; VASP, vasodilator-stimulated phosphoprotein

* Corresponding author.

E-mail address: vera.ralevic@nottingham.ac.uk (V. Ralevic).

¹ Current address: Platelet Solutions Ltd., BioCity Nottingham, Pennyfoot Street, Nottingham NG1 1GF, UK.

<https://doi.org/10.1016/j.vph.2017.12.063>

Received 5 April 2017; Received in revised form 4 December 2017; Accepted 12 December 2017

1537-1891/ © 2018 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

pyrimidine and sugar nucleotides. There are eight members of the P2Y family, namely P2Y₁, P2Y₂, P2Y₄, P2Y₆, P2Y₁₁, P2Y₁₂, P2Y₁₃ and P2Y₁₄ receptors, with the P2Y₁₄ receptor being the most recently described [4,9]. The P2Y₁₄ receptor is a G_{i/o} protein-coupled receptor activated by UDP-sugars including UDP-glucose, UDP-galactose, UDP-glucuronic acid, and UDP-N-acetylglucosamine [4,5,16,17,23,44–46]. The P2Y₁₄ receptor is also activated by UDP, and by MRS2690, a more selective agonist that has a 7-fold greater potency at the P2Y₁₄ receptor than UDP-glucose [31]. Quantitative analysis has revealed that the P2Y₁₄ protein is expressed in many regions including the brain, adipose tissue, lung, spleen, heart and certain immune and inflammatory cells [9].

There is evidence for the expression of P2Y₁₄ receptor transcript and protein in vascular smooth muscle cells [5,19,38], but relatively little is known about the functional expression of this receptor in the cardiovascular system. Until recently, the lack of availability of P2Y₁₄ receptor antagonists made it difficult to determine a role for the P2Y₁₄ receptor. A novel non-nucleotide compound, 4-((piperidin-4-yl)-phenyl)-7-(4-(trifluoromethyl)-phenyl)-2-naphthoic acid (PPTN), was reported as a high-affinity competitive antagonist at the P2Y₁₄ receptor [43]. PPTN inhibited UDP-glucose and MRS2690 mediated mast cell degranulation, and UDP-glucose mediated chemotaxis of neutrophils [6,16,17]. We recently used PPTN to show, for the first time, that UDP-glucose mediates vasoconstriction through activation of P2Y₁₄ receptors in porcine pancreatic arteries [5]. UDP-glucose mediates contraction in mouse coronary arteries, and the response was retained in P2Y₂ knockouts, however since neither a P2Y₁₄ selective agonist nor antagonist were employed the identity of the receptor involved requires confirmation [21]. The aims of the current study were to determine whether UDP-sugars can modulate contractility of the porcine coronary artery and to characterize the P2 receptor(s) involved.

Since the P2Y₁₄ receptor is G_{i/o} protein-coupled, this makes its investigation difficult in isolated tissues where cAMP levels are normally low. Accordingly, we used forskolin to stimulate adenylyl cyclase and raise cAMP levels, a method described by us [5] and others [42] previously to investigate vascular G_{i/o} protein-coupled receptors, and on this background we were able to investigate the pharmacology of the responses to UDP-glucose and other P2Y₁₄ receptor agonists in the porcine coronary artery. In this study we report the functional expression of a contractile P2Y₁₄ receptor sensitive to UDP-sugars in the porcine coronary artery. The contractile response was mimicked by the selective P2Y₁₄ agonist, MRS2690, and the response was blocked by the selective P2Y₁₄ receptor antagonist PPTN. Measurement of VASP phosphorylation, and of contractile responses in the presence of forskolin, indicated that UDP-glucose inhibits cAMP levels, consistent with the known G_{i/o} protein-coupling of the P2Y₁₄ receptor.

2. Methods

2.1. Tissue preparation

Hearts were obtained from pigs (either sex, ~6 months old, weight ~50 kg) and were transported on ice from a local abattoir. The size and arterial system of the porcine heart are very similar to that of humans [49]. Porcine left circumflex coronary arteries were dissected out and stored overnight at 4 °C in oxygenated (95% O₂, 5% CO₂) Krebs-Henseleit buffer (mM): sodium chloride 118; potassium chloride 4.8; magnesium sulphate 1.1; sodium hydrogen carbonate 25; potassium hydrogen phosphate 1.2; glucose 12; calcium chloride 1.25. A fine dissection was carried out the following day. Four adjacent 5 mm ring segments were cut from each vessel and mounted onto wires for isometric tension recording (Grass FT03 transducers) in warmed (37 °C) and gassed (95% O₂, 5% CO₂) Krebs-Henseleit buffer in organ baths. In some arteries the endothelium was removed by gentle rolling of the segment with fine-tipped forceps inserted into the lumen [41]. The success of the treatment was evaluated by testing responsiveness to an endothelium-dependent vasodilator, substance P.

2.2. Responses in the porcine coronary artery

Rings were put under an initial tension of 10 g [41] and left for ~45 min to equilibrate, after which viability was assessed by eliciting contractions with 60 mM KCl. After washout and equilibration (30 min) cumulative concentration response curves to UDP-glucose (0.1 μM–1 mM), MRS2690 (1 nM–10 μM), UDP-glucuronic acid (0.1 μM–1 mM), and UDP-N-acetylglucosamine (0.1 μM–1 mM) were constructed at basal tone.

2.3. Forskolin elevation of cAMP

In some experiments, arteries were preconstricted with U46619 (5–10 nM), a thromboxane A₂ mimetic, to elicit contraction of 35–75% of that to KCl. Forskolin (0.1 μM; activates adenylyl cyclase) was used to reverse the U46619-mediated contraction. The combination of forskolin and U46619 has previously been used to uncover α₂-adrenoceptor-mediated contractions in porcine ear artery [42]. When vessels had reached a stable tone (30–45 min), cumulative-concentration response curves to UDP-glucose, MRS2690, UDP-glucuronic acid and UDP-N-acetylglucosamine were constructed. In separate experiments, PPTN (1 μM) was added 10 min prior to addition of U46619. Cumulative concentration-response curves to UDP-glucose, MRS2690 and UTP were constructed in the absence and presence of PPTN (1 μM). The effects of MRS2578 (10 μM, a P2Y₆ receptor antagonist), and the P2 receptor antagonists PPADS (10 μM) and suramin (100 μM), were also investigated. In some experiments sodium nitroprusside (0.1 μM; activates guanylyl cyclase) was used instead of forskolin, together with U46619, to investigate effects on contractility to UDP-glucose. In some experiments UDP-glucose was treated with apyrase (5 U/ml, 30 min) before use and apyrase (5 U/ml) was also added to the organ baths prior to U46619 and forskolin additions.

2.4. VASP phosphorylation assay

Porcine coronary arteries were set up in organ baths as described above. After addition of U46619 (5–10 nM) and forskolin (0.1 μM), vessels were left to relax to basal tone. These were control vessels, which were paired with vessels where either 100 μM or 1 mM UDP-glucose, or 10 μM MRS2690, were added in the absence and presence of PPTN (1 μM). After the contractions had reached a maximum, vessels were removed, put into lysis buffer, and frozen in dry ice. Tissues were homogenized and the supernatant was separated by centrifugation (3000 RCF, 10 min). VASP-phosphorylation in the tissues was measured by flow cytometry using a cytometric bead assay where VASP in the samples was captured onto beads using monoclonal anti-VASP (IE273) followed by detection using FITC conjugated anti-VASP (5C6 anti-VASP pSer 157, Acris Antibodies, Germany) [27].

2.5. Preparation of platelets

Venous blood was obtained from 7 healthy volunteers (from School of Life Sciences, University of Nottingham). Ethics was approved by Thrombosis and Haemostasis Research Group, University of Nottingham. Blood samples from healthy volunteers were taken in accordance with local ethical permission and the declaration of Helsinki. Blood was immediately put into collecting tubes containing EDTA (4 mM) to prevent blood clotting. For platelet preparation, platelet rich plasma was obtained by centrifugation at 1100 RPM (15 min) followed by a 2000 RPM (20 min) centrifugation step to obtain a platelet pellet. Platelets were resuspended in sterile saline solution (0.9% sodium chloride) before centrifuging at 1600 RMP (15 min). The pellet was resuspended in 2 ml of saline. Cells were counted on Sysmex KX-21 Haematology analyser (Kobe, Japan). Cells were between 78.3 and 92.9% pure (contamination with red and white blood cells).

2.6. Platelet addition to porcine coronary arteries

Porcine coronary arteries were prepared as previously described. In some vessels the P2Y₁₄ receptor antagonist PPTN (1 μ M) was added 10 min prior to U46619 addition. After a stable tone had been established, isolated platelets were added to the vessels; the final platelet concentration in the bath was approximately 24,000–30,000/ μ L. In other experiments, L-NAME (100 μ M) was added to porcine coronary arteries and left for 10 min before adding PPTN (1 μ M). After pre-contraction with U46619, the isolated platelets were added.

2.7. Materials

Acetylcholine, apyrase (Grade III), histamine, uridine 5'-diphosphoglucose disodium salt (UDP-glucose), uridine 5'-diphosphoglucuronic acid trisodium salt (UDP-glucuronic acid), uridine 5'-diphospho-N-acetylglucosamine sodium salt (UDP-N-acetylglucosamine), uridine 5'-triphosphate trisodium salt hydrate (UTP), 7-beta-acetoxy-8,13-epoxy-1-alpha, 6-beta, 9-alpha-trihydroxyabund-14-en-11-one; coleonol (adenylyl cyclase activator: forskolin), and N ω -nitro-L-arginine methyl ester hydrochloride (L-NAME) were all obtained from Sigma Aldrich. Diphosphoric acid 1- α -D-glucopyranosyl ester 2-[(4'-methylthio)uridin-5"-yl] ester disodium salt (MRS2690) and N,N"-1,4-Butanediybis[N'-(3-isothiocyanatophenyl)thiourea] (MRS2578) were obtained from Tocris Biosciences Ltd. PPTN was a gift from Merck Frosst Centre for Therapeutic Research.

2.8. Statistical analysis

Data were fitted using non-linear regression and statistical analyses were performed using GraphPad Prism 6.0 software. R_{max} is the maximal response and logEC₅₀ is the log of the molar concentration of agonist required to generate 50% of the R_{max}. Differences between R_{max} and logEC₅₀ values were compared using the extra sum of squares F test, by comparing the extra sum of squares that resulted from the analysis with separate R_{max} or logEC₅₀ values; where out-of-sample extrapolation occurred all parameters were selected to determine if one curve adequately fit all data sets (Prism 6). Individual R_{max} and logEC₅₀ values were also determined after curve fitting of data for each artery segment. These parameters were compared using one- or two-way ANOVA with Tukey's multiple comparisons post hoc test to determine which groups the differences were between when > 2 groups were being compared. Data from VASP-phosphorylation experiments were analysed using one-way ANOVA with Tukey's post test or Student's *t*-test. Data from platelet experiments were analysed by two-way ANOVA with repeated measures, with Fisher's least significant difference post hoc test. *P* < 0.05 was considered statistically significant. All data are presented as mean \pm S.E.M. *n* signifies the number of animals.

3. Results

3.1. P2Y₁₄ receptor agonists constrict coronary arteries

The P2Y₁₄ receptor is activated by various sugar nucleotides, including UDP-glucose, UDP-glucuronic acid and UDP-N-acetylglucosamine [4] and, more potently, by the synthetic analogue MRS2690 [31]. To identify and characterize the P2Y₁₄ receptor in the porcine coronary artery, cumulative concentration-response curves to UDP-glucose (0.1 μ M–1 mM) and MRS2690 (1 nM–10 μ M) were initially constructed at basal tone. The agonists were found to produce concentration-dependent contractions (Fig. 1). MRS2690 was found to be more potent than UDP-glucose; R_{max} and EC₅₀ values are reported in Table 1. Since the P2Y₁₄ receptor is a G_{i/o} protein-coupled receptor mediating inhibition of adenylyl cyclase activity and hence reducing cAMP production [44–46], arteries were pre-exposed to forskolin to enhance intracellular cAMP levels prior to constructing the

concentration-response curves. U46619 (5–10 nM), a thromboxane A₂ mimetic, was additionally present in these experiments to maintain a stable basal tone (since forskolin induces vasorelaxation). Under these conditions, contractile responses to UDP-glucose and MRS2690 were enhanced as shown in Fig. 1. When the experiment was carried out using U46619 together with sodium nitroprusside (SNP, activator of guanylyl cyclase) instead of forskolin, no augmentation of the responses to UDP-glucose was observed, consistent with a specific involvement of cAMP (Fig. 1e). There was a significant difference in the R_{max} values for UDP-glucose between the groups (*P* < 0.001; extra sum of squares F-test). Analysis of the data (ANOVA with Tukey's post hoc test) showed that there was a significant increase in the R_{max} in the presence of forskolin plus U46619 (R_{max} 1.69 \pm 0.24 g, *n* = 6) compared to the controls (R_{max} 0.35 \pm 0.09 g, *n* = 6; *P* < 0.001) and compared to the presence of SNP plus U46619 (R_{max} 0.57 \pm 0.1 g, *n* = 12; *P* < 0.001). The logEC₅₀ values for UDP-glucose were similar under the different experimental conditions: control –5.4 \pm 0.41 (*n* = 6), forskolin plus U46619 –4.66 \pm 0.14 (*n* = 6) and SNP plus U46619 –5.03 \pm 0.08 (*n* = 12).

The combination of forskolin plus U46619 failed to augment contractile responses to histamine and acetylcholine (Fig. 2), which cause vasoconstriction of porcine coronary arteries primarily via G_{q/11} coupled H₁ histamine receptors [22] and M₃ muscarinic receptors [13]. R_{max} values for ACh were similar in the absence and presence of forskolin plus U46619: 8.94 \pm 1.07 g (*n* = 5) and 9.53 \pm 1.13 g (*n* = 6) respectively. logEC₅₀ values were also similar in the absence and presence of forskolin plus U46619: –6.50 \pm 0.08 (*n* = 5) and –6.57 \pm 0.1 (*n* = 6), respectively (Fig. 2a). R_{max} values for histamine were similar in the absence and presence of forskolin plus U46619: 8.84 \pm 0.52 g (*n* = 4) and 9.71 \pm 0.8 g (*n* = 4), respectively. There was, however, a small but significant difference in the logEC₅₀ values for histamine in the presence of forskolin plus U46619 compared to control conditions: –5.15 \pm 0.1 (*n* = 4) and –5.45 \pm 0.07 (*n* = 4), respectively (*P* < 0.01, extra sum of squares F-test) (Fig. 2b), indicating a small reduction in the potency of histamine in the presence of forskolin plus U46619.

UDP-glucuronic acid and UDP-N-acetylglucosamine (0.1 μ M–1 mM) also produced concentration-dependent contractions of the porcine coronary arteries (Fig. 3; Table 1). All four of the P2Y₁₄ receptor agonists (UDP-glucose, UDP-glucuronic acid, UDP-N-acetylglucosamine and MRS2690) produced greater contractions in tissues exposed to forskolin (and U46619) (Fig. 3b) than in control tissues (Fig. 3a), although analysis of the R_{max} values showed that this difference was only statistically significant for UDP-glucose and UDP-glucuronic acid (ANOVA, Table 1). An increase in contractility under these experimental conditions is consistent with an involvement of G_{i/o} protein coupled receptors and inhibition of cAMP. There was a significant difference between the logEC₅₀ values of the agonists (*P* < 0.001), but their R_{max} values were similar (*P* > 0.05) (extra sum of squares F-test). Analysis of the logEC₅₀ values (ANOVA with Tukey's post test) showed that MRS2690 was significantly more potent than the other UDP-sugars (Table 1). UMP (0.1 μ M–1 mM) did not cause contraction of the porcine coronary arteries (tested in the presence of forskolin and U46619; data not shown).

Removal of the endothelium had no significant effect on the contractile response to UDP-glucose in the porcine coronary artery, showing that this was mediated by direct actions on the smooth muscle (Fig. 4a); R_{max} values were 2.96 \pm 0.22 g (*n* = 12) and 3.12 \pm 0.36 g (*n* = 11) in endothelium-intact and -denuded arteries, respectively, and logEC₅₀ values were –4.25 \pm 0.13 (*n* = 12) and –4.34 \pm 0.14 (*n* = 11), respectively. Substance P (10 nM) was used to confirm the presence of the endothelium in the porcine coronary arteries: in endothelium-intact arteries relaxation was 41 \pm 6.4% (*n* = 12); in endothelium-denuded arteries substance P did not cause a relaxation (*n* = 11).

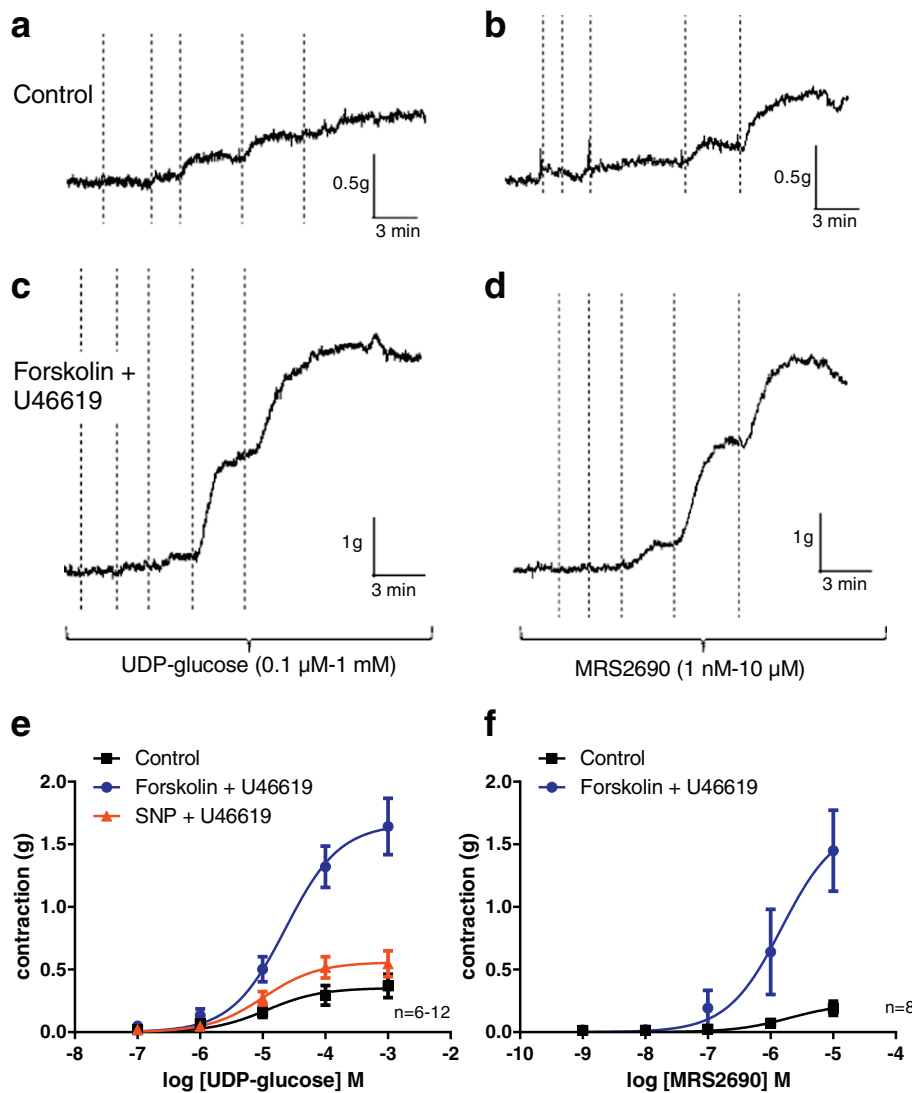


Fig. 1. UDP-glucose and the selective P2Y₁₄ receptor agonist MRS2690 mediate contraction of porcine coronary arteries. Representative traces showing concentration-dependent contractile responses of porcine coronary arteries to UDP-glucose (a, c) and MRS2690 (b, d) at basal tone in the absence (Control) and presence of forskolin (0.1 μM) plus U46619. Lines on traces represent addition of cumulative concentrations of agonists. The mean data ± S.E.M. are of experiments from 6 to 12 different pigs (e, f). For UDP-glucose (e) the R_{max} was significantly different between the groups ($P < 0.001$, extra sum of squares F test) with no difference in $\log EC_{50}$ values. For MRS2690 (f) the curves were significantly different ($P < 0.001$, extra sum of squares F test).

3.2. Effects of suramin and PPADS, P2 receptor antagonists, on contractile responses to UDP-glucose in coronary arteries

The P2 receptor antagonist suramin (100 μM) reduced the maximal contractile response to UDP-glucose (obtained in the presence of

forskolin plus U46619) from 2.94 ± 0.45 g ($n = 7$) to 2.12 ± 0.29 g ($n = 8$) ($P < 0.01$, extra sum of squares F test) (Fig. 4b). The $\log EC_{50}$ values were not significantly different at -4.74 ± 0.04 ($n = 7$) and -4.50 ± 0.07 ($n = 8$), in the absence and presence of suramin respectively. These data are consistent with an involvement of P2 receptors

Table 1

Contractile responses of the porcine coronary artery to UDP-sugars. Shown are the EC_{50} ($-\log M$) and R_{max} values.

| Agonist | $EC_{50} \pm S.E.M. (-\log M)$ | | $R_{max} \pm S.E.M. (contraction in g)$ | |
|-------------------------|---------------------------------|------------------------------------|---|-----------------------|
| | Control | Forskolin (+ U46619) | Control | Forskolin (+ U46619) |
| UDP-glucose | 4.89 ± 0.21 $n = 12$ | 4.66 ± 0.14 $n = 6$ | 0.37 ± 0.06 | $1.69 \pm 0.24^{\#}$ |
| UDP-glucuronic acid | 4.80 ± 0.27 $n = 7$ | 3.63 ± 0.21 $n = 7$ | 0.42 ± 0.04 | $2.26 \pm 0.51^{###}$ |
| UDP-N-acetylglucosamine | 4.86 ± 0.23 $n = 7$ | 4.27 ± 0.11 $n = 7$ | 0.49 ± 0.10 | 1.72 ± 0.31 |
| MRS2690 | $6.30 \pm 0.45^{**}$ $n = 4$ | $5.71 \pm 0.16^{\phi}$ $n = 12$ | 0.48 ± 0.14 | 1.77 ± 0.32 |

Contractile response curves were investigated in arteries under control conditions (Control) and in arteries to which forskolin had been added to stimulate adenylyl cyclase (with U46619 added to maintain a stable tone; see Methods) (forskolin). $**P < 0.01$ for MRS2690 $-\log EC_{50}$ compared with $-\log EC_{50}$ s of UDP-glucose, UDP-glucuronic acid and UDP-N-acetylglucosamine in controls (two way ANOVA with Tukey's post test). $\phi P < 0.05$, $P < 0.001$, $P < 0.001$ for MRS2690 $-\log EC_{50}$ compared with $-\log EC_{50}$ s of UDP-glucose, UDP-glucuronic acid and UDP-N-acetylglucosamine respectively, in the presence of forskolin (two-way ANOVA with Tukey's post test). Agonist R_{max} values in the presence of forskolin were significantly greater compared to corresponding R_{max} values obtained under control conditions for both UDP-glucose and UDP-glucuronic acid; $^{\#}P < 0.5$, $^{###}P < 0.001$ (two-way ANOVA with Tukey's post test). Data are mean ± S.E.M.

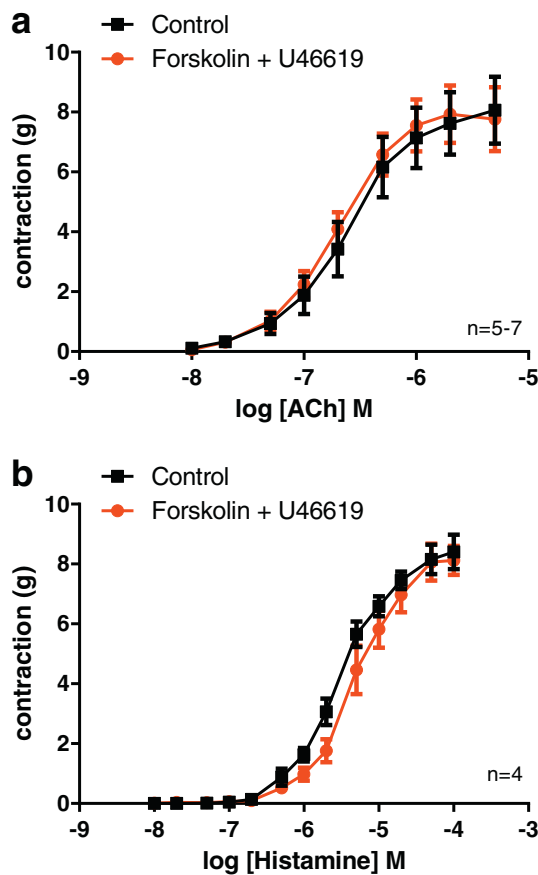


Fig. 2. Forskolin plus U46619 do not augment contractions to acetylcholine and histamine in porcine coronary arteries. Cumulative concentration-response curves to acetylcholine (ACh) (a) and histamine (b), under control conditions and the effect of forskolin (0.1 μ M) plus U46619. There was no significant difference in the R_{max} values for either compound, or in the $\log EC_{50}$ values for ACh, but there was a small but significant difference in $\log EC_{50}$ values for histamine, indicating a slightly reduced potency in the presence of forskolin plus U46619 ($P < 0.01$, extra sum of squares F-test). Data are mean \pm S.E.M.

as mediators of the contractile response to UDP-glucose, but do not indicate which P2 receptor subtype(s) are involved. In contrast, PPADS (10 μ M) augmented the contractile response to UDP-glucose (obtained in the presence of forskolin plus U46619) (Fig. 4c); the R_{max} was significantly greater in the presence of PPADS at 2.42 ± 0.23 g ($n = 6$) than in the control at 2.03 ± 0.16 g ($n = 6$) ($P < 0.01$, extra sum of squares F test) and the sensitivity of the response to UDP-glucose was also increased in the presence of PPADS, $\log EC_{50} = -5.77 \pm 0.05$ ($n = 6$) compared to the control, $\log EC_{50} = -4.81 \pm 0.08$ ($n = 6$) ($P < 0.001$, extra sum of squares F test).

3.3. PPTN, a selective P2Y₁₄ receptor antagonist, blocks responses to MRS2690, a selective P2Y₁₄ receptor agonist, in coronary arteries

In order to investigate the possible involvement of the P2Y₁₄ receptor in the porcine coronary artery, PPTN was used to examine contractile responses to the P2Y₁₄ receptor agonists UDP-glucose and MRS2690. PPTN at 1 μ M acts selectively to inhibit the P2Y₁₄ receptor with no effect at the other P2Y receptor subtypes [6]. In the presence of PPTN (1 μ M), the contractile responses to UDP-glucose were attenuated and there was a significant difference between the response curves ($P < 0.01$, extra sum of squares F-test) (Fig. 5a). The contractions to MRS2690 were also attenuated by PPTN with a significant difference between the response curves ($P < 0.001$, extra sum of squares F-test) (Fig. 5b).

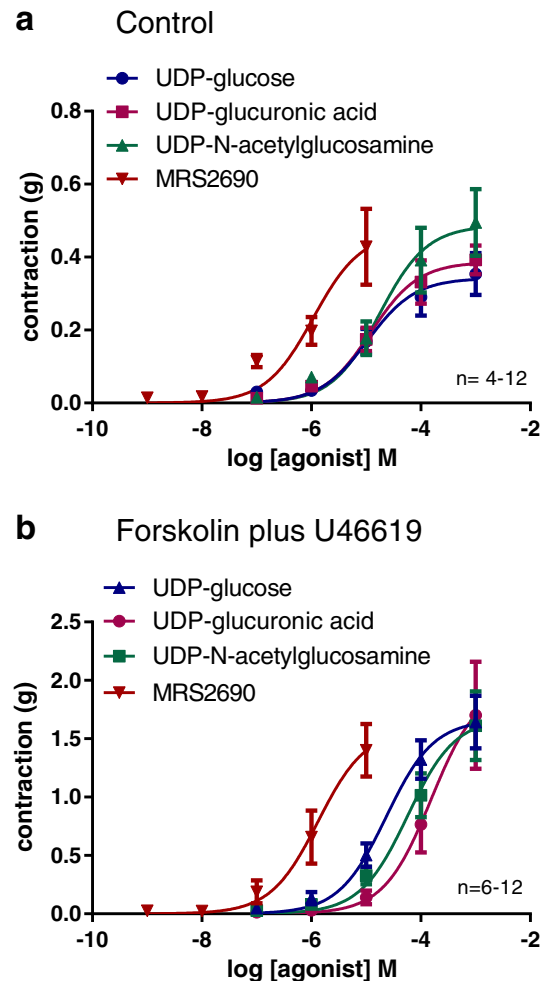


Fig. 3. UDP-sugars mediate contraction of porcine coronary arteries and the responses are augmented by forskolin. Cumulative concentration-response curves to P2Y₁₄ receptor agonists in the porcine coronary artery. UDP-glucose, UDP-glucuronic acid, UDP-N-acetylglucosamine and the P2Y₁₄ receptor agonist MRS2690 were investigated in isolated porcine coronary arteries under control conditions (a) and in arteries to which forskolin (0.1 μ M) and U46619 had been added to elevate cAMP and maintain tone, respectively (b). Note the different y axes scales in panels a and b. There was a significant difference in the agonist $\log EC_{50}$ values in the presence of forskolin plus U46619 ($P < 0.001$, extra sum of squares F test) (see also Table 1). Data are mean \pm S.E.M.

UTP is an agonist at P2Y₂ and P2Y₄ receptors, and elicits contraction of porcine coronary arteries through activation of these receptors [41]. UTP does not have any effect at the P2Y₁₄ receptor [9]. To determine the selectivity of action of PPTN versus other P2Y receptors in the PCA, namely P2Y₂ and P2Y₄ receptors, the effect of PPTN on contractile responses to UTP was investigated. PPTN had no significant effect on the contractile responses to UTP (Fig. 5c).

3.4. MRS2578, a selective P2Y₆ receptor antagonist, does not block responses to MRS2690 and UDP-glucose in coronary arteries

P2Y₆ receptors are present in the porcine coronary artery and, upon activation by their endogenous ligand UDP, elicit vasoconstriction [41]. We investigated whether UDP-glucose and MRS2690 cause a P2Y₆ receptor mediated contractile response in porcine coronary arteries. In addition, in order to determine whether the contractions to UDP-glucose involved UDP, either present as a contaminant or following release from the tissue, the UDP-glucose solution was treated with the nucleotidase apyrase (5 U/ml, 30 min) and apyrase was also added to the organ baths (5 U/ml, prior to addition of forskolin and U46619). We found that neither MRS2578 (10 μ M), a selective P2Y₆ receptor

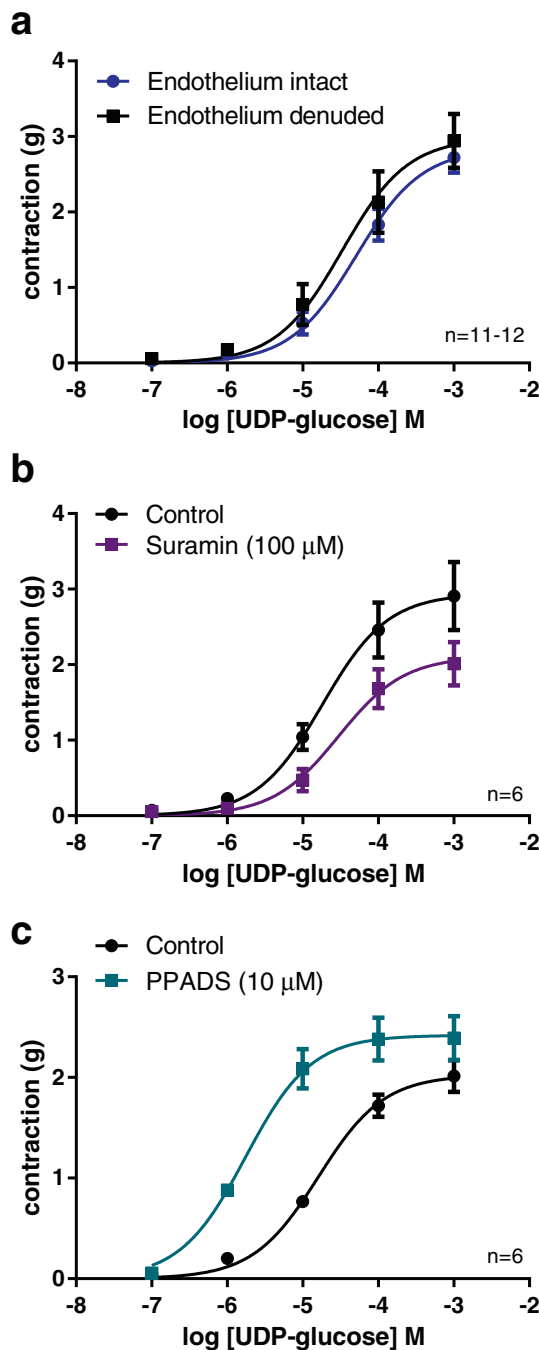


Fig. 4. Contraction to UDP-glucose in porcine coronary arteries is endothelium-independent and attenuated by suramin but augmented by PPADS. Concentration-response curves to UDP-glucose in endothelium-intact and endothelium-denuded arteries in the presence of forskolin plus U46619 (a). Effect of the P2 receptor antagonists suramin (b) and PPADS (c) on contractile responses to UDP-glucose in porcine coronary arteries in the presence of forskolin plus U46619. UDP-glucose R_{max} values were significantly different in the presence and absence of suramin ($P < 0.01$) and both R_{max} and $\log EC_{50}$ values were significantly different in the presence and absence of PPADS ($P < 0.01$ and $P < 0.001$) (extra sum of squares F test). Data are mean \pm S.E.M.

antagonist, nor apyrase, had any significant effect on contractile responses induced by UDP-glucose, in coronary arteries in the presence of forskolin and U46619 (Fig. 6a). UDP-glucose R_{max} values were: 2.93 ± 0.49 g ($n = 7$), 2.92 ± 0.70 g ($n = 6$), 2.68 ± 0.46 g ($n = 6$) and 2.81 ± 0.54 g ($n = 5$), in controls and in the presence of

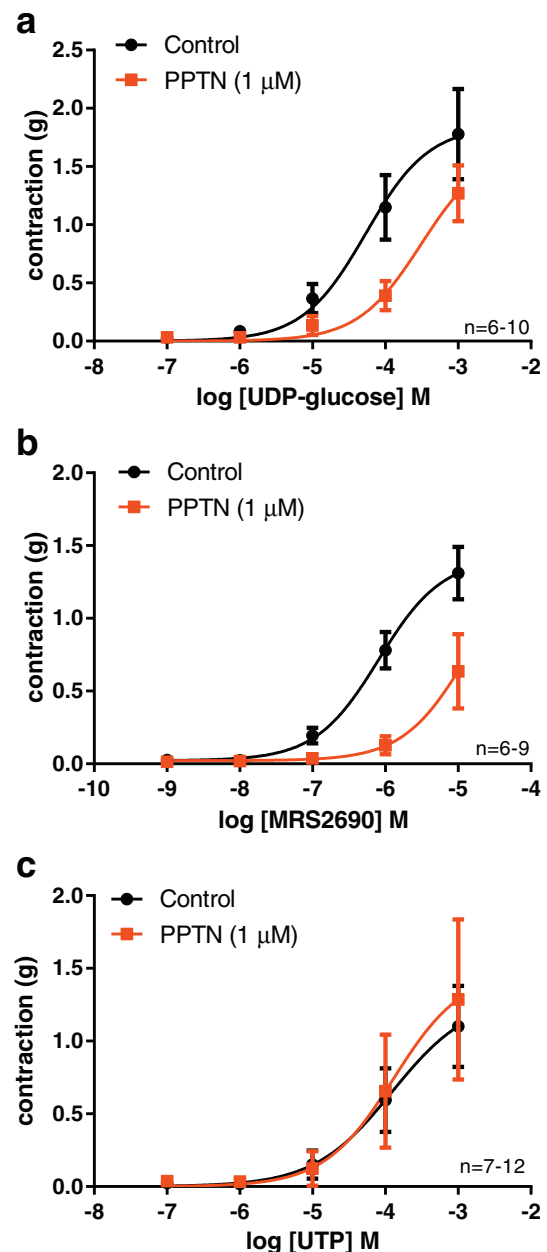


Fig. 5. Contractile responses to UDP-glucose and the selective P2Y₁₄ receptor agonist MRS2690 are attenuated by the selective P2Y₁₄ receptor antagonist PPTN. The effect of PPTN (1 μ M) on contractile responses to UDP-glucose (a), MRS2690 (b), and UTP (c) in porcine coronary arteries in the presence of forskolin (0.1 μ M) and U46619. Analysis of the curves showed that these were significantly different in the absence and presence of PPTN for UDP-glucose ($P < 0.01$) and MRS2690 ($P < 0.001$) (extra sum of squares F test). In contrast, there was no significant difference in the responses to UTP. Data are mean \pm S.E.M.

MRS2578, apyrase and MRS2578 plus apyrase, respectively; the corresponding $\log EC_{50}$ values were -5.26 ± 0.11 , -5.45 ± 0.28 , -5.48 ± 0.16 , and -5.58 ± 0.16 . MRS2578 also had no significant effect on contractions mediated by MRS2690 in coronary arteries in the presence of forskolin and U46619, in line with the selectivity of this compound for P2Y₁₄ receptors; R_{max} values were 3.38 ± 0.66 g ($n = 8$) in controls and 2.71 ± 0.54 g ($n = 10$) in the presence of MRS2578 and the corresponding $\log EC_{50}$ values were -6.76 ± 0.22 and -6.70 ± 0.13 (Fig. 6b).

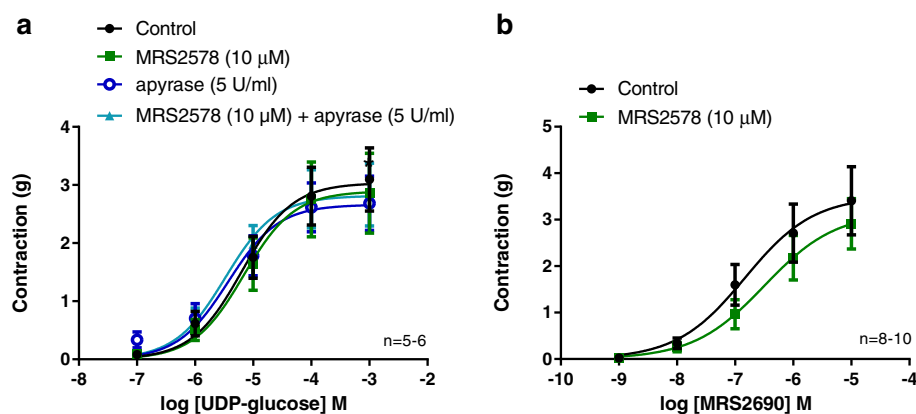


Fig. 6. Contractile responses to UDP-glucose and MRS2690 are unaffected by the selective P2Y₆ receptor antagonist MRS2578. Effect of MRS2578, on concentration-response curves to (a) UDP-glucose and (b) MRS2690 in porcine coronary arteries. UDP-glucose and MRS2690 caused concentration-dependent contractions in the presence of forskolin and U46619. Contractions to UDP-glucose were unaffected by MRS2578 or by prior incubation of the UDP-glucose solution and tissues with apyrase (5 U/ml, > 30 min) (a). Contractions to MRS2690 were also unaffected by MRS2578. Data are mean \pm S.E.M.

3.5. P2Y₁₄ receptor agonists decrease VASP-phosphorylation in coronary arteries

Increases in intracellular cAMP levels lead to the phosphorylation of vasodilator-stimulated phosphoprotein (VASP) via cAMP-dependent protein kinase A (PKA) and other kinases. Thus VASP-phosphorylation (VASP-P) can be used to measure levels of cAMP. In the present study, forskolin was used to increase cAMP levels in porcine coronary arteries. Forskolin alone (0.1 μM), an activator of adenylate cyclase, significantly increased VASP-P levels ($P < 0.001$, ANOVA with Tukey's post test) (Fig. 7a). UDP-glucose (100 μM) and MRS2690 (10 μM) significantly decreased the forskolin-elevated levels of VASP-P, to levels that were similar to those measured in control tissues ($P < 0.05$, ANOVA with Tukey's post test) (Fig. 6a). PPTN (1 μM) was able to partially restore levels of VASP-P that were decreased by UDP-glucose and MRS2690 in porcine coronary arteries (Fig. 7b). PPTN alone (1 μM) had no significant effect on VASP-P levels in forskolin exposed porcine coronary arteries (Fig. 7c). Together these results suggest that P2Y₁₄ receptor activation couples to the inhibition of adenylate cyclase activity, hence reducing levels of cAMP and VASP-P in porcine coronary arteries in vitro.

3.6. Effect of platelets in porcine coronary arteries and the effect of PPTN

Isolated platelets (24,700–30,000 cells/μl) added to coronary artery segments that had been preconstricted with U46619 caused a significant relaxation ($P < 0.01$, two-way ANOVA) which reversed over time (Fig. 8a–c). PPTN (1 μM) did not significantly change the mean relaxation response induced by the platelets from 3 donors (Fig. 8a–c). L-NAME (100 μM), a nitric oxide synthase inhibitor, was added to vessels to remove the platelet-induced relaxations, and in its presence platelets caused predominantly contraction ($P < 0.001$, two-way ANOVA) (Fig. 8d–g). PPTN had no significant effect on the mean platelet-induced contractions in the presence of L-NAME in 3 donors (Fig. 8e–g), although for one donor the platelet-induced contractile response was augmented (two-way ANOVA) (Fig. 8d).

4. Discussion

We have provided the first evidence for the functional expression of the P2Y₁₄ purine receptor in the porcine coronary vasculature. Vasoconstriction was produced by UDP-glucose, UDP-glucuronic acid, UDP-N-acetylglucosamine and MRS2690, all of which are known P2Y₁₄ receptor agonists [4,9], and MRS2690 was more potent than the UDP-sugars, consistent with previous characterization of MRS2690 as a potent and selective P2Y₁₄ receptor agonist [31]. Responses to UDP-glucose and MRS2690 were selectively antagonized by PPTN, a competitive antagonist at the P2Y₁₄ receptor [6,43], but were unaffected by the P2Y₆ receptor antagonist MRS2578.

Forskolin is routinely added to cultured and dissociated cells to elevate cAMP levels in order to reveal the function of G_{i/o} protein-coupled receptors, and although less commonly used in intact tissue it has been used to uncover vasoconstrictive α₂-adrenoceptors in the porcine ear artery [42] and P2Y₁₄ receptors in the porcine pancreatic artery [5]. Vasoconstrictive responses to the endogenous and synthetic P2Y₁₄ receptor agonists were increased in vessels in the presence of forskolin, but not when the forskolin was substituted with sodium nitroprusside (increases cGMP). Moreover, the combination of U46619 and forskolin had no effect on contractions to histamine and acetylcholine, which in porcine coronary arteries are mediated through activation of H₁ histamine receptors [22] and M₃ muscarinic receptors [13] respectively; both receptors are coupled to G_{q/11} proteins and therefore do not inhibit adenylate cyclase activity, suggesting that forskolin treatment was not augmenting responses to the vasoconstrictors non-selectively. These observations are consistent with the involvement of G_{i/o} protein-coupled P2Y₁₄ receptors causing inhibition of adenylate cyclase activity and a reduction in cAMP. Pertussis toxin treatment (which blocks G_{i/o} protein-coupling) inhibits UDP-glucose binding to P2Y₁₄ receptors [4,9], but we were not able to use pertussis toxin successfully in our studies, possibly because of difficulties with it permeating the multilayered tissue. Physiologically, elevations in cAMP levels might occur following activation of G_s protein-coupled β-adrenoceptors and A_{2A} and A_{2B} adenosine receptors and concomitant UDP-sugar activation of P2Y₁₄ receptors would be expected to limit vasodilatation, and other functions, mediated by these receptors.

The selective P2Y₁₄ receptor agonist MRS2690 was more potent than UDP-glucose, UDP-glucuronic acid and UDP-N-acetylglucosamine, consistent with the literature [16,28]. Contractions to MRS2690 were evident at concentrations equal to and below 10 μM, and at 10 μM MRS2690 is inactive at P2Y₂ receptors [30]. PPTN, which inhibits P2Y₁₄ receptors but not other P2Y receptors [6,43], blocked contractile responses of the porcine coronary artery to MRS2690 and the responses were virtually abolished at 1 μM MRS2690. Contractile responses to UTP (agonist at P2Y₂ and P2Y₄ receptors) were unaffected by PPTN at a concentration that inhibited responses to MRS2690. Moreover, the contractile response to MRS2690 was unaffected by the P2Y₆ receptor antagonist MRS2578 [37]. These results indicate that P2Y₁₄ receptors are functionally expressed in porcine coronary arteries, along with P2Y₂, P2Y₄ and P2Y₆ receptors, for which the principal endogenous ligands are UTP and ATP (P2Y₂), UTP (P2Y₄) and UDP (P2Y₆) [41].

With regard to the endogenous ligands, contractile responses to UDP-glucose were mimicked by UDP-glucuronic acid and UDP-N-acetylglucosamine, consistent with an involvement of P2Y₁₄ receptors. This is unlike UDP-glucose responses in N9 microglia which were not mimicked by UDP-galactose and thus were suggested to be independent of P2Y₁₄ receptors [7]. Moreover, responses to UDP-glucose were blocked by PPTN, indicating actions at P2Y₁₄ receptors, and consistent with a role for UDP sugars as extracellular signalling molecules. Contractile

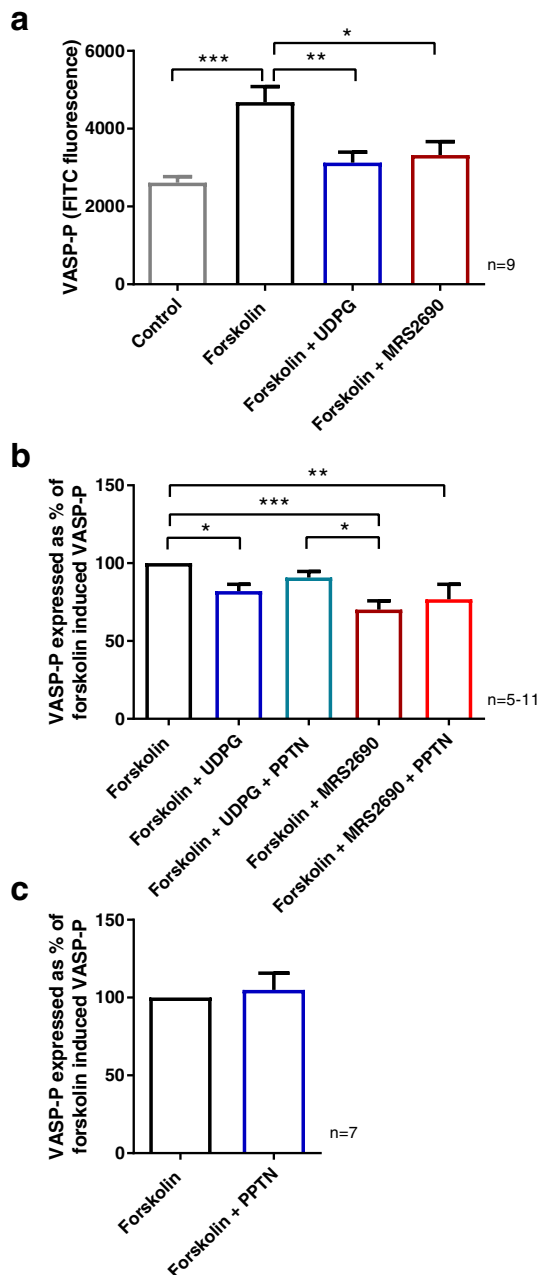


Fig. 7. UDP-glucose-induced VASP-phosphorylation is blocked by the P2Y₁₄ receptor antagonist PPTN. Levels of VASP phosphorylation (VASP-P) in porcine coronary arteries at basal tone, in the absence (Control) and presence of forskolin and the effect of UDP-glucose (UDPG), MRS2690 (P2Y₁₄ receptor agonist) and PPTN (P2Y₁₄ receptor antagonist). (a) Levels of VASP-P in control conditions and in the presence of forskolin, forskolin + UDP-glucose (100 μ M), and forskolin + MRS2690 (10 μ M). (b) Effect of UDP-glucose (1 mM) and MRS2690 (10 μ M) on forskolin-induced VASP phosphorylation in porcine coronary arteries in the absence and presence of PPTN (1 μ M). (c) Effect of PPTN (1 μ M) on forskolin-induced VASP-P. VASP-phosphorylation was measured using flow cytometry. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, ANOVA with Tukey's post test. PPTN alone had no effect on VASP-P (Student's t -test). Data are mean \pm S.E.M.

responses to UDP-glucose were also attenuated by the non-selective P2 receptor antagonist suramin. The non-selective P2 receptor antagonist PPADS augmented contractile responses to UDP-glucose, as reported by us previously in porcine pancreatic arteries [5]; the mechanism remains to be determined but does not appear to involve the ectonucleotidase inhibitory activity of PPADS [10,20] and potentiation of responses to contaminant nucleotides, since our experiments with apyrase indicate that nucleotides are not present as major contaminants of UDP-glucose,

as reported by others [47]. There is evidence for interactions between cAMP and Ca²⁺ signalling pathways such that an increase in intracellular Ca²⁺ inhibits cAMP accumulation [11]. There is also evidence that PPADS antagonises IP₃-induced intracellular Ca²⁺ mobilization [50] and that it elevates basal cAMP levels [40]. Thus, it is possible that a PPADS-induced elevation of cAMP levels (further to the elevation caused by forskolin), possibly involving a reduction in intracellular Ca²⁺, leads to its facilitation of contractile responses to UDP-glucose.

Actions at multiple P2Y receptor targets is common among purine/pyrimidine nucleotides [8] but responses to UDP-glucose were not inhibited by MRS2578 indicating a lack of involvement of vasocontractile P2Y₆ receptors [41]. Another possible target is the P2Y₂ receptor, at which UDP-glucose is known to act, albeit weakly [30], and which is present in the porcine coronary artery [41]. In coronary and basilar arteries from P2Y₂ knockout mice the response to UDP-glucose was slightly, but not significantly, smaller than that in arteries from wild type mice [21]. UMP and uridine are known not to contribute to P2Y₁₄ receptor activation [9], and we found that UMP did not contract porcine coronary arteries (unpublished observations).

Since endothelium removal had no effect on contractions to UDP-glucose, the P2Y₁₄ receptors mediating this response are likely to be located on the coronary artery smooth muscle. We have shown that the P2Y₁₄ receptor is located on both the smooth muscle and endothelium of porcine pancreatic arteries [5], reinforcing that P2 receptor expression can vary with blood vessel as well as species [8]. Expression of mRNA encoding P2Y₁₄ was shown in rat and mouse aortic smooth muscle [19,29]. Meister et al. [38] also reported P2Y₁₄ expression in blood vessels, but mainly in veins. In immunoblotting experiments for P2Y₁₄ receptor protein we showed a band at 41 kDa, which is close to the predicted molecular weight of 38 kDa for the P2Y₁₄ receptor, and an additional, stronger band was seen at 61 kDa (Supplemental data). Similar sized bands (34 kDa and 56 kDa) were reported for P2Y₁₄ in C6 glioma cells, and evidence was provided of a contribution of glycosylation to the higher molecular weight band [33]. However, the specificity of commercially available antibodies is a concern (see [18]) and validation of the P2Y₁₄ receptor antibody is needed to understand whether these bands are the P2Y₁₄ receptor.

In parallel studies, VASP-P was measured as an indicator of cAMP accumulation in the porcine coronary artery. Forskolin significantly enhanced VASP-P, and UDP-glucose and MRS2690 inhibited the forskolin-induced increase in VASP-P. Moreover PPTN was able to partially reverse the UDP-glucose and MRS2690 mediated inhibition of forskolin-induced VASP-P, consistent with an involvement of P2Y₁₄ receptors mediating a reduction in cAMP levels. PPTN was similarly shown to antagonise UDP-glucose-mediated inhibition of adenylyl cyclase activity in C6 glioma cells expressing P2Y₁₄ receptors [6]. UDP-glucose also inhibited forskolin-stimulated cAMP accumulation in T-lymphocytes, neutrophils and human astrocytoma cells [44–46]. Overall, the data suggest that as in other tissues, the P2Y₁₄ receptor in the porcine coronary artery is G_{i/o} protein-coupled.

The present findings are important because there is evidence that UDP-glucose may be a more stable extracellular signalling molecule than ATP and UTP, both of which have been shown to be released from heart during coronary hypoxia/ischaemia [14,15,24]. Release of UDP-glucose has been shown from several cell types (including human astrocytoma cells, Calu-3 airway epithelial cells, COS-7, CHO-K1 and C6 glioma cells) and, significantly, unlike ATP levels, the stimulated levels of UDP-glucose remained elevated for up to 3 h [36]. Similarly, there was negligible metabolism of extracellular UDP-glucose in neutrophil suspensions at 1 h, at which time ATP had been completely hydrolysed by neutrophil NTPDase1 (nucleotide triphosphate diphosphohydrolase-1), which hydrolyses extracellular nucleotides but not UDP-sugars [47]. This has important implications for P2Y₁₄ receptor signalling during cardiac hypoxia, ischaemia, inflammation, or cell damage, during which UDP-glucose may be released within the coronary vasculature.

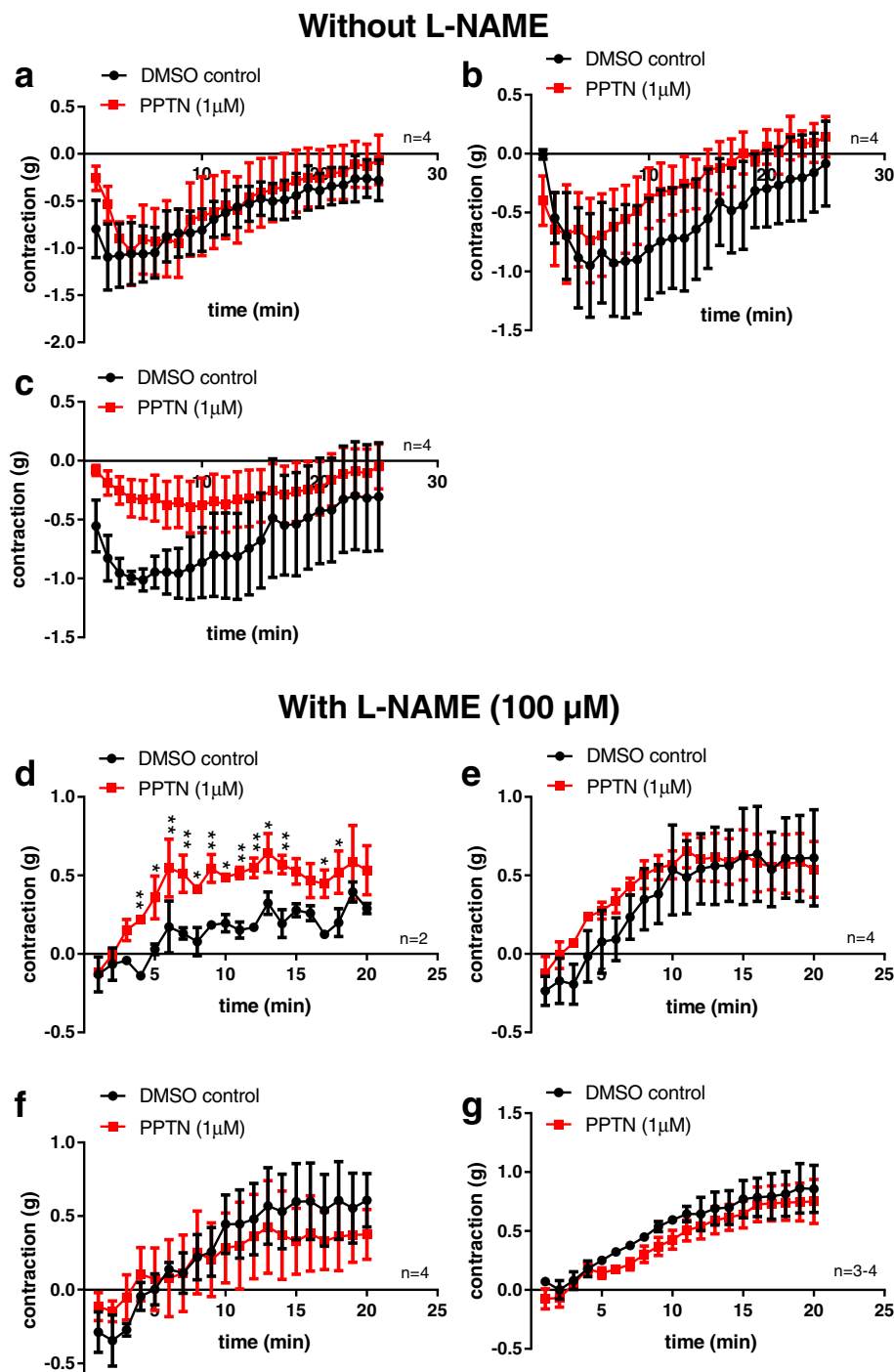


Fig. 8. Effect of platelets and the P2Y₁₄ receptor antagonist PPTN, on porcine coronary artery contractility. Platelets isolated from 7 healthy adult human donors were added to U46619-precontracted porcine coronary arteries in the absence (a–c) and presence (d–g) of L-NAME (100 μ M), a nitric oxide synthase inhibitor, in the absence and presence of PPTN (1 μ M) (P2Y₁₄ receptor antagonist). In the absence of L-NAME the platelets elicited vasorelaxation. In the presence of L-NAME platelets elicited vasoconstriction. *n* indicates the number of different pigs from which coronary artery segments were obtained. **P* < 0.05; ***P* < 0.01 for comparison of responses in the absence and presence of PPTN (two way ANOVA). Data are mean \pm S.E.M.

UDP-glucose levels of $\sim 90 \mu\text{M}$ were determined in heart extracts [34] and are in line with the effective concentrations of UDP-glucose producing vasoconstriction of the coronary arteries in the present study. Such levels might be found following exocytotic release of UDP-glucose or upon cell damage in the heart or following hypoxia, ischaemia or inflammation. Other potential sources of UDP-sugars in the cardiovascular system include platelets, leukocytes and endothelial cells. It follows that antagonists at P2Y₁₄ receptors may have therapeutic potential in ischaemic heart disease.

UDP-sugars including UDP-glucose, UDP-galactose, UDP-glucuronic acid and UDP-*N*-acetylglucosamine have been measured in platelets [51] and we were interested to see if UDP-sugars are released from activated platelets to mediate contraction of coronary arteries. In the

porcine coronary arteries the addition of platelets from the human donors produced a relaxation, which was unaffected by PPTN. In canine coronary arteries, aggregating platelets cause an endothelium-dependent relaxation which is largely due to the actions of ADP and ATP released from the platelets and this relaxation was reduced in vessels treated with apyrase [25,26]. In endothelium denuded canine coronary arteries, platelets elicit vasoconstriction that does not occur in endothelium intact vessels [26,48]. Thus, we used an inhibitor of nitric oxide synthase, L-NAME, to uncover the contractile response to the platelets. This is the first study to use L-NAME to block platelet-induced relaxations in vascular tissue and thus to show a specific involvement of NO. The porcine coronary arteries contracted when platelets were added in the presence of L-NAME, likely due to actions of contractile

mediators including 5-HT and thromboxane A₂ released from the activated platelets [26], but PPTN had no significant effect on this response in 3 of the donors and augmented the response in one, suggesting that P2Y₁₄ receptors do not contribute significantly under the conditions of the present study. The effects of the UDP-sugars may be small in comparison to those of the other components released from platelets. High levels of P2Y₁₄ receptor mRNA have been detected in megakaryocytic cells and platelets [39]. Although P2Y₁₄ receptors have been characterised in platelets using immunoblotting analysis, UDP-glucose and MRS2690 did not have any effect on a number of human platelet functions [12].

In summary, UDP-glucose mediates contraction of the porcine coronary artery via P2Y₁₄ receptors. The P2Y₁₄ receptor is coupled negatively to adenylyl cyclase since UDP-glucose and the selective P2Y₁₄ receptor agonist MRS2690 decreased cAMP accumulation as determined by lower levels of VASP-P; these decreased levels of VASP-P could partially be restored by PPTN, a selective P2Y₁₄ receptor antagonist. P2Y₁₄ receptors may be involved in the control of coronary arterial blood flow and may, therefore, be possible targets for the treatment of ischaemic heart disease.

Funding

This work was supported by the British Heart Foundation [Grant number FS/12/7/29359].

Conflict of interest

None declared.

Acknowledgments

We would like to thank Frosst Centre for Therapeutic Research for gifting us with the PPTN compound.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.vph.2017.12.063>.

References

- [1] Z.S.B. Abbas, N. Dovlatova, S.C. Fox, W.R. Dunn, V. Ralevic, Effect of a novel P2Y₁₄ receptor antagonist on the porcine coronary artery, *Proc. Br. Pharmacol. Soc.* 11 (3) (2013), <http://www.pa2online.org/abstracts/vol11issue3abst192p.pdf> (Abstract).
- [2] Z. Abbas, W. Dunn, V. Ralevic, Effect of forskolin on contractile responses to the P2Y₁₄ receptor agonists UDP-glucose and MRS2690 in porcine coronary arteries, *Proc. Br. Pharmacol. Soc.* 10 (4) (2012), <http://www.pa2online.org/abstracts/vol10issue4abst057p.pdf> (Abstract).
- [3] Z. Abbas, L. Latif, V. Ralevic, Effects of P2Y₁₄ receptor agonists, UDP-glucose and MRS2690, on porcine coronary artery contractility, *Proc. Br. Pharmacol. Soc.* 9 (3) (2011), <http://www.pa2online.org/abstracts/vol9issue3abst051p.pdf> (Abstract).
- [4] M.P. Abbracchio, J.M. Boeynaems, E.A. Barnard, J.L. Boyer, C. Kennedy, M.T. Miras-Portugal, B.F. King, C. Gachet, K.A. Jacobson, G.A. Weisman, G. Burnstock, Characterization of the UDP-glucose receptor (re-named here the P2Y₁₄ receptor) adds diversity to the P2Y receptor family, *Trends Pharmacol. Sci.* 24 (2) (2003) 52–55.
- [5] M. Alsaqati, M.L. Latif, S.L.F. Chan, V. Ralevic, Novel vasocontractile role of the P2Y₁₄ receptor: characterisation of its signalling in porcine isolated pancreatic arteries, *Br. J. Pharmacol.* 171 (2014) 701–713.
- [6] M.O. Barrett, J.I. Sesma, C.B. Ball, P.S. Jayasekara, K.A. Jacobson, E.R. Lazarowski, T.K. Harden, A selective high-affinity antagonist of the P2Y₁₄ receptor inhibits UDP-glucose-stimulated chemotaxis of human neutrophils, *Mol. Pharmacol.* 84 (1) (2013) 41–49.
- [7] V.M. Brautigam, G.R. Dubyak, J.M. Crain, J.J. Watters, The inflammatory effects of UDP-glucose in N9 microglia are not mediated by P2Y₁₄ receptor activation, *Purinergic Signal.* 4 (1) (2008) 73–78.
- [8] G. Burnstock, V. Ralevic, Purinergic signalling and blood vessels in health and disease, *Pharmacol. Rev.* 66 (2014) 102–192.
- [9] J.K. Chambers, L.E. Macdonald, H.M. Sarau, R.S. Ames, K. Freeman, J.J. Foley, Y. Zhu, M.M. McLaughlin, P. Murdock, L. McMillan, J. Trill, A. Swift, N. Aiyar, P. Taylor, L. Vawter, S. Naheed, P. Szekeres, G. Hervieu, C. Scott, J.M. Watson, A.J. Murphy, E. Duzic, C. Klein, D.J. Bergsma, S. Wilson, G.P. Livi, A G protein-coupled receptor for UDP-glucose, *J. Biol. Chem.* 275 (15) (2000) 10767–10771.
- [10] B.C. Chen, C.M. Lee, W.W. Lin, Inhibition of ecto-ATPase by PPADS, suramin and reactive blue in endothelial cells, C6 glioma cells and RAW 264.7 macrophages, *Br. J. Pharmacol.* 119 (8) (1996) 1628–1634.
- [11] M. DeBernardi, T. Sekl, G. Brooker, Inhibition of cAMP accumulation by intracellular calcium mobilization in C6-2B cells stably transfected with substance K receptor cDNA, *Proc. Natl. Acad. Sci. U. S. A.* 88 (1991) 9257–9261.
- [12] N. Dovlatova, Y.D. Wijeyeratne, S.C. Fox, P. Manolopoulos, A.J. Johnson, A.E. White, M.L. Latif, V. Ralevic, S. Heptinstall, Detection of P2Y₁₄ protein in platelets and investigation of the role of P2Y₁₄ in platelet function in comparison with the EP₃ receptor, *Thromb. Haemost.* 100 (2008) 261–270.
- [13] M. Entzeroth, H.N. Doods, N. Mayer, Characterization of porcine coronary muscarinic receptors, *Naunyn. Arch. Pharmacol.* 341 (5) (1990) 432–438.
- [14] D. Erlinge, J. Harnek, C. van Heusden, G. Olivecrona, S. Jern, E. Lazarowski, Uridine triphosphate (UTP) is released during cardiac ischemia, *Int. J. Cardiol.* 100 (3) (2005) 427–433.
- [15] T. Forrester, C.A. Williams, Release of adenosine triphosphate from isolated adult heart cells in response to hypoxia, *J. Physiol.* 268 (2) (1977) 371–390.
- [16] Z.G. Gao, Y. Ding, K.A. Jacobson, UDP-glucose acting at P2Y₁₄ receptors is a mediator of mast cell degranulation, *Biochem. Pharmacol.* 79 (6) (2010) 873–879.
- [17] Z.G. Gao, Q. Wei, M.P. Jayasekara, K.A. Jacobson, The role of P2Y₁₄ and other P2Y receptors in degranulation of human LAD2 mast cells, *Purinergic Signal.* 9 (1) (2013) 31–40.
- [18] G. Griffiths, J.M. Lucocq, Antibodies for immunolabeling by light and electron microscopy: not for the faint hearted, *Histochem. Cell Biol.* 142 (4) (2014) 347–360.
- [19] S. Govindan, E.J. Taylor, C.W. Taylor, Ca²⁺ signalling by P2Y receptors in cultured rat aortic smooth muscle cells, *Br. J. Pharmacol.* 160 (8) (2010) 1953–1962.
- [20] B. Grobbs, K. Anciaux, D. Roymans, C. Stefan, M. Bollen, E.L. Esmans, H. Slegers, An ecto-nucleotide pyrophosphatase is one of the main enzymes involved in the extracellular metabolism of ATP in rat C6 glioma, *J. Neurochem.* 72 (2) (1999) 826–834.
- [21] K.A. Haanes, L. Eddvinsson, Characterization of the contractile P2Y₁₄ receptor in mouse coronary and cerebral arteries, *FEBS Lett.* 588 (17) (2014) 2936–2943.
- [22] M. Hagen, I. Paegelow, Histamine H1- and H2-receptors in coronary arteries of pigs, *Agents Actions* 9 (3) (1979) 253–256.
- [23] T.K. Harden, J.I. Sesma, I.P. Fricks, E.R. Lazarowski, Signalling and pharmacological properties of the P2Y receptor, *Acta Physiol.* 199 (2) (2010) 149–160.
- [24] A.M. Hopwood, J. Lincoln, K.A. Kirkpatrick, G. Burnstock, Adenosine 5'-triphosphate, adenosine and endothelium-derived relaxing factor in hypoxic vasodilation of the heart, *Eur. J. Pharmacol.* 165 (2–3) (1989) 323–326.
- [25] D.S. Houston, J.T. Shepherd, P.M. Vanhoutte, Adenine nucleotides, serotonin, and endothelium-dependent relaxations to platelets, *Am. J. Phys.* 248 (1985) H389–395.
- [26] D.S. Houston, J.T. Shepherd, P.M. Vanhoutte, Aggregating human platelets cause direct contraction and endothelium-dependent relaxation of isolated canine coronary arteries. Role of serotonin, thromboxane A₂, and adenine nucleotides, *J. Clin. Invest.* 78 (1986) 539–544.
- [27] D. Iyu, J.R. Glenn, A.E. White, A. Johnson, S. Heptinstall, S.C. Fox, The role of prostanoic receptors in mediating the effects of PGE₃ on human platelet function, *Thromb. Haemost.* 107 (4) (2012) 797–799.
- [28] K.A. Jacobson, A.A. Ivanov, S. de Castro, T.K. Harden, H. Ko, Development of selective agonists and antagonists of P2Y receptors, *Purinergic Signal.* 5 (1) (2009) 75–89.
- [29] G. Kauffenstein, A. Drouin, N. Thorin-Trescases, H. Bachelard, B. Robaye, P. D'Orleans-Juste, F. Marceau, E. Thorin, J. Sévigny, NTPDase1 (CD39) controls nucleotide-dependent vasoconstriction in mouse, *Cardiovasc. Res.* 85 (1) (2010) 204–213.
- [30] H. Ko, A. Das, R.L. Carter, I.P. Flocks, Y. Zhou, A.A. Ivanov, A. Melman, B.V. Joshi, P. Kováč, J. Hajdúch, K.L. Kirk, T.K. Harden, Molecular recognition in the P2Y₁₄ receptor: probing the structurally permissive terminal sugar moiety of uridine-5'-diphosphoglucose, *Bioorg. Med. Chem.* 17 (14) (2009) 5298–5311.
- [31] H. Ko, I. Fricks, A.A. Ivanov, T.K. Harden, K.A. Jacobson, Structure-activity relationship of uridine 5'-diphosphoglucose analogues as agonists of the human P2Y₁₄ receptor, *J. Med. Chem.* 50 (9) (2007) 2030–2039.
- [32] S. Kook, J. Cho, S.B. Lee, B.C. Lee, The nucleotide sugar UDP-glucose mobilizes long-term repopulating primitive hematopoietic cells, *J. Clin. Invest.* 123 (8) (2013) 3420–3435.
- [33] P. Krzeminski, P. Pomorski, J. Baranska, The P2Y₁₄ receptor activity in glioma C6 cells, *Eur. J. Pharmacol.* 594 (1–3) (2008) 49–54.
- [34] M.R. Laughlin, W.A. Petit, J.M. Dizon, R.G. Schulman, E.J. Barrett, NMR measurements of in vivo myocardial glycogen metabolism, *J. Biol. Chem.* 263 (1988) 2285–2291.
- [35] E.R. Lazarowski, Vesicular and conductive mechanisms of nucleotide release, *Purinergic Signal.* 8 (2012) 359–373.
- [36] E.R. Lazarowski, D.A. Shea, R.C. Boucher, T.K. Harden, Release of cellular UDP-glucose as a potential extracellular signaling molecule, *Mol. Pharmacol.* 63 (5) (2003) 1190–1197.
- [37] L.K. Mamedova, B.V. Joshi, Z.G. Gao, I. von Kugelgen, K.A. Jacobson, Diisothiocyanate derivatives as potent, insurmountable antagonists of P2Y₆ nucleotide receptors, *Biochem. Pharmacol.* 67 (2004) 1763–1770.
- [38] J. Meister, D. le Duc, A. Ricken, R. Burkhardt, J. Thiery, H. Pfannkuche, T. Plote, J. Grosse, T. Schöneberg, A. Schulz, The G protein-coupled receptor P2Y₁₄ influences insulin release and smooth muscle function in mice, *J. Biol. Chem.* 289 (34) (2014) 23353–23366.
- [39] D.J. Moore, P.R. Murdock, J.M. Watson, R.L. Faull, H.J. Waldvogel, P.G. Szekeres,

- S. Wilson, K.B. Freeman, P.C. Emson, GPR105, a novel $G_{i/o}$ -coupled UDP-glucose receptor expressed on brain glia and peripheral immune cells, is regulated by immunologic challenge: possible role in neuroimmune function, *Brain Res. Mol. Brain Res.* 118 (2003) 10–23.
- [40] S. Ohkubo, K. Kumazawa, K. Sagawa, J. Kimura, I. Matsuoka, β , γ -methylene ATP-induced cAMP formation in C6Bu-1 cells: involvement of local metabolism and subsequent stimulation of adenosine A_{2B} receptor, *J. Neurochem.* 76 (2001) 872–880.
- [41] S.J. Rayment, M.L. Latif, V. Ralevic, S.P. Alexander, Evidence for the expression of multiple uracil nucleotide-stimulated P2 receptors coupled to smooth muscle contraction in porcine isolated arteries, *Br. J. Pharmacol.* 150 (5) (2007) 604–612.
- [42] R.E. Roberts, A.E. Tomlinson, D.A. Kendall, V.G. Wilson, α_2 -adrenoceptor-mediated contractions of the porcine isolated ear artery: evidence for a cyclic AMP-dependent and a cyclic AMP-independent mechanism, *Br. J. Pharmacol.* 124 (6) (1998) 1107–1114.
- [43] J. Robichaud, J.F. Fournier, S. Gagne, J.Y. Gauthier, M. Hamel, Y. Han, M. Hénault, S. Kargman, J.-F. Levesque, Y. Mamane, J. Mancini, N. Morin, E. Mulrooney, J. Wu, W.C. Black, Applying the pro-drug approach to afford highly bioavailable antagonists of P2Y₁₄, *Bioorg. Med. Chem. Lett.* 21 (14) (2011) 4366–4368.
- [44] M. Scrivens, J.M. Dickenson, Pharmacological effects mediated by UDP-glucose that are independent of P2Y₁₄ receptor expression, *Pharmacol. Res.* 51 (6) (2005) 533–538.
- [45] M. Scrivens, J.M. Dickenson, Functional expression of the P2Y₁₄ receptor in murine T-lymphocytes, *Br. J. Pharmacol.* 146 (3) (2005) 435–444.
- [46] M. Scrivens, J.M. Dickenson, Functional expression of the P2Y₁₄ receptor in human neutrophils, *Eur. J. Pharmacol.* 543 (1–3) (2006) 166–173.
- [47] J.I. Sesma, S.M. Kreda, N. Steinckwich-Besancon, H. Dang, R. García-Mata, T.K. Harden, E.R. Lazarowski, The UDP-sugar-sensing P2Y₁₄ receptor promotes Rho-mediated signaling and chemotaxis in human neutrophils, *Am. J. Phys. Cell Phys.* 303 (5) (2012) C490–C498.
- [48] H. Shimokawa, P.M. Vanhoutte, Angiographic demonstration of hyperconstriction induced by serotonin and aggregating platelets in porcine coronary arteries with regenerated endothelium, *J. Am. Coll. Cardiol.* 17 (1991) 1197–1202.
- [49] M.M. Swindle, A. Makin, A.J. Herron, F.J. Clubb, K.S. Frazier, Swine as models in biochemical research and toxicology testing, *Vet. Pathol.* 49 (2) (2011) 344–356.
- [50] P. Vigne, P. Pacaud, V. Urbach, E. Feolde, J.P. Breittmayer, C. Frelin, The effect of PPADS as an antagonist of inositol (1,4,5)trisphosphate induced intracellular calcium mobilization, *Br. J. Pharmacol.* 119 (1996) 360–364.
- [51] H.H. Wandall, V. Rumjantseva, A.L. Sorensen, S. Patel-Hett, E.C. Josefsson, E.P. Bennett, J.E. Italiano Jr., H. Clausen, J.H. Hartwig, K.M. Hoffmeister, The origin and function of platelet glycosyltransferases, *Blood* 120 (2012) 626–635.